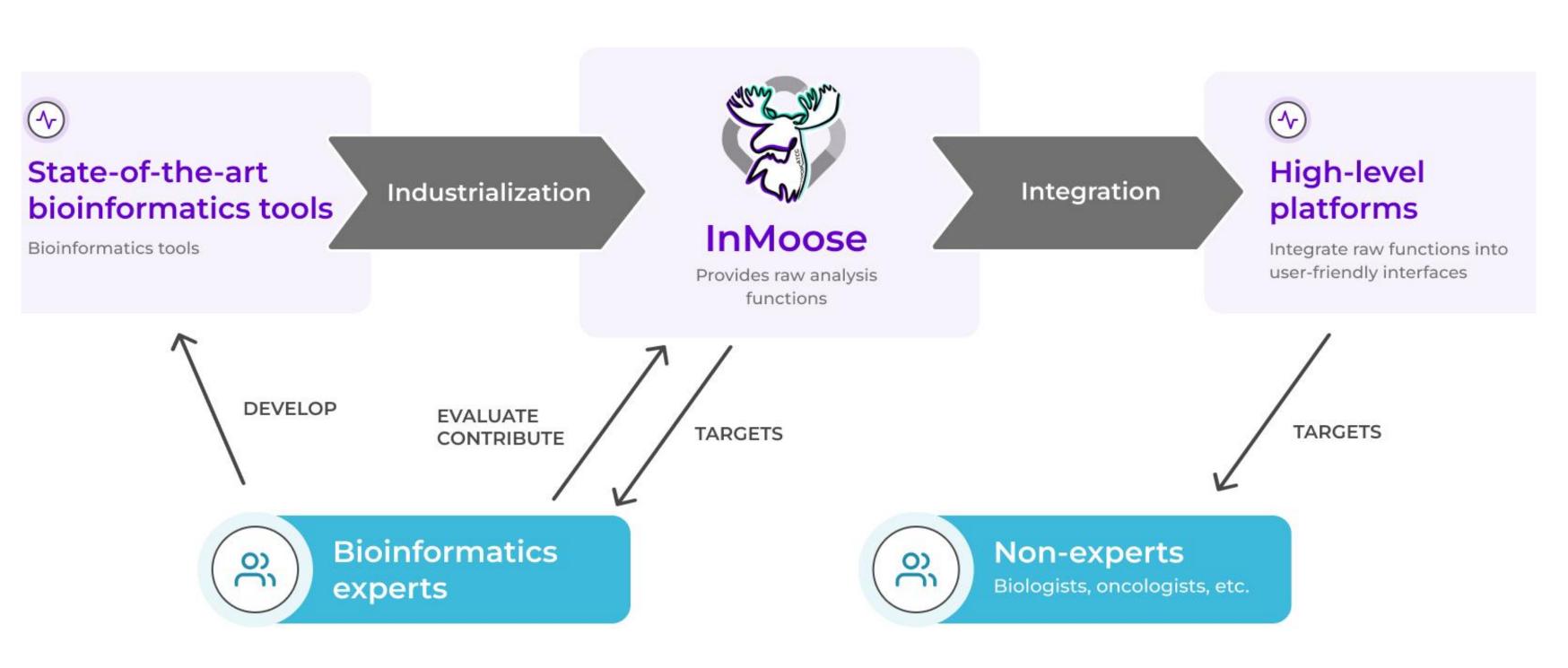


Introducing InMoose

An Integrated Multi-omic Open Source Package for Python Analyses





Introduction

Exponentially increasing amount of omic data exacerbates the need for high-quality, high-performance and highly interoperable computational tools for cancer research.

Our goal is to industrialize tried-and-tested, high-quality tools, by making them more efficient and more interoperable.

Open Source

- •InMoose is open source (GPL3 License).
- •Open source is our way to **bind with the community**.
- •Open source is a great way to achieve **quality and performance**:
- anyone can audit the code (evaluation)
- anyone can contribute to the code (improvement)
- Open source fosters interoperability:
- easier to interface with other tools
- •Open source is a prerequisite for **open science**.
- pyComBat GitHub repository:
- 10+ external issues, 42 stars, 18 forks
- •pyComBat preprint: 16 citations

Approach

Our method is to port existing tools to Python, and integrate them in a single package.

As a general-purpose, mainstream language, Python offers several perks:

- easy integration into large-scale frameworks (e.g. web platform)
 - ⇒ **versatility**, user-friendliness, wide target audience
- widespread language
 - ⇒ accessibility (e.g. cross-disciplinary collaboration)
- trendy in bioinformatics
 - ⇒ harmonizing ecosystem, momentum
- porting = opportunity window for improvement
 - ⇒ functionality, **performance** (e.g. ComBat-Seq)

First tools ported:

- batch effect correction (ComBat, ComBat-Seq)
- differential expression analysis (DESeq2)

Our approach aims to foster a larger collaborative effort to build and grow a consistent state-of-the-art Python ecosystem for cancer bioinformatics.

Results

Our approach is to faithfully port the code from R/C++ to Python/C++, to preserve the quality of the ported tools.

We assess the quality of our ports by replicating results of the original tools.

- Microarray batch effect correction
 - Results are reproduced with a mean relative difference of 2.5e⁻⁷% (CI95%: [3.4e⁻¹¹,1.6e⁻⁶]).
 - InMoose runs almost 4x faster than the original R implementation.
- RNAseq batch effect correction
 - Results are reproduced with a mean relative difference of 5.4e⁻⁸% (first non-zero quantile 0.9995).
 - o InMoose runs more than **4x faster** than the original R implementation.

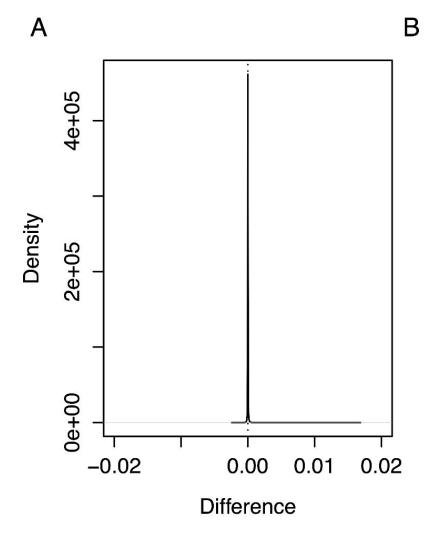
Performance of InMoose vs. Combat and ComBat-Seq.

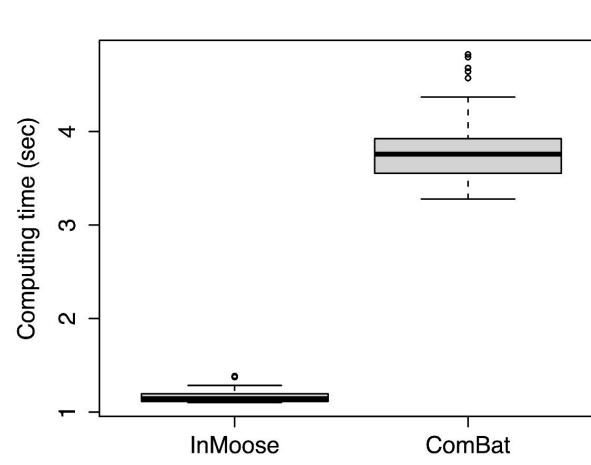
A - Distribution of the relative differences between the expression matrices corrected for batch effects, respectively by ComBat and InMoose (parametric version). The vertical dotted line corresponds to zero.

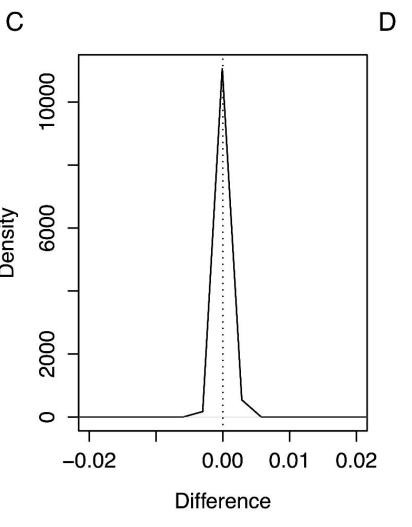
B - Computation time in seconds for InMoose and ComBat for the parametric method.

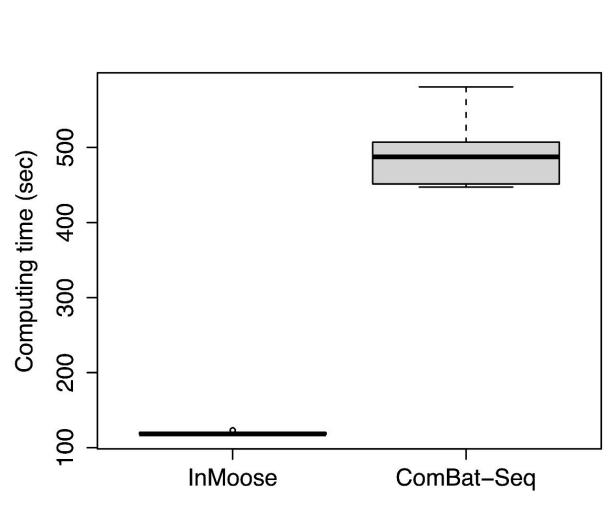
C - Distribution of the relative differences between the expression matrices corrected for batch effects, respectively by ComBat-Seq and InMoose. The vertical dotted line corresponds to zero.

D - Computation time in seconds for InMoose and ComBat-Seq.









References

- 1. Johnson, W. E., Li, C., & Rabinovic, A. (2007). *Adjusting batch effects in microarray expression data using empirical Bayes methods*. Biostatistics, 8(1), 118–127.
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- 4. Love, M. I., Huber, W., & Anders, S. (2014). *Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2*. Genome Biology, 15(12), 550.
- 5. Leek, J. T., Johnson, W. E., Parker, H. S., Fertig, E. J., Jaffe, A. E., Zhang, Y., Storey, J. D., & Torres L. C. (2022). sva: Surrogate Variable Analysis. R package version 3.46.0.

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Check it out!

- •QR code to PyPI page
- •Call to contribution and feedback



